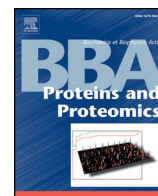




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Review

Q1 Perspective on computational and structural aspects of kinase discovery from IPK2014[☆]

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ABSTRACT

Recent advances in understanding the activity and selectivity of kinase inhibitors and their relationships to protein structure are presented. Conformational selection in kinases is studied from empirical, data-driven and simulation approaches. Ligand binding and its affinity are, in many cases, determined by the predetermined active and inactive conformation of kinases. Binding affinity and selectivity predictions highlight the current state of the art and advances in computational chemistry as it applies to kinase inhibitor discovery. Kinome wide inhibitor profiling and cell panel profiling lead to a better understanding of selectivity and allow for target validation and patient tailoring hypotheses. This article is part of a Special Issue entitled: Inhibitors of Protein Kinases.

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1. Introduction

Understanding the relationship of kinase targets in normal and disease states and their modulation by inhibitors stands at a crossroads in the discovery and delivery of new medicines. Overcoming key challenges such as target selection, pathway modulation, compound prioritization, understanding toxicity, biomarker selection and patient tailoring is key to the design of better treatments. Computational sciences, including bio, chemo, and structural informatics are increasingly indispensable in kinase discovery. Chemical and structural informatics streamline complexity, finding patterns in large data sets and generating testable hypotheses for experimentation. Several talks at the conference reported analyses of large datasets of structural and activity data of

many compounds tested with many kinases. Alexander Baumann, Richard Engh and Thibault Varin described platforms for experimental activity profiling of compounds across large kinase panels, and computational methods to understand patterns of cross-reactivity across compound, kinase, and cell line dimensions. Eric Martin used arrays of kinase predictive models to estimate inhibition profiles when experimental data are incomplete or lacking. Henrik Moebitz and Stefan Knapp evaluated large databases of X-ray structures and compound activities to relate protein conformational states to binding affinity. Benoit Roux used physics-based molecular dynamics simulations rather than informatics to understanding relative energies of DFG-in and DFG-out kinase conformations. Valerio Berdini employed a chemistry-based approach to the conformation problem, building up ligands to DFG-in and DFG-out conformations from fragment-based starting points. Another aspect of kinase computation correlates chemical similarity with kinase potency to predict activity in lieu of or in advance or experiments. Thibault Varin, Eric Martin and Jens Meiler described ligand-based kinase inhibition and selectivity models, and their impact on drug discovery projects. In addition to their predictive, explanatory aspects, computational sciences play an increasing role in experiment design spanning a range from target hypotheses to compound design. This

Abbreviations: Abl, Abelson murine leukemia; MD, Molecular dynamics; Melk, Maternal embryonic leucine zipper kinase; Kit, v-kit Hardy–Zuckerman 4 feline sarcoma viral oncogene homolog; JNK, c-Jun N-terminal kinase; PLK, Polo like kinase; CDK, Cyclin dependent kinase; ERK, Extracellular-signal-regulated kinase; Pdb, Protein data bank; NMR, Nuclear magnetic resonance.

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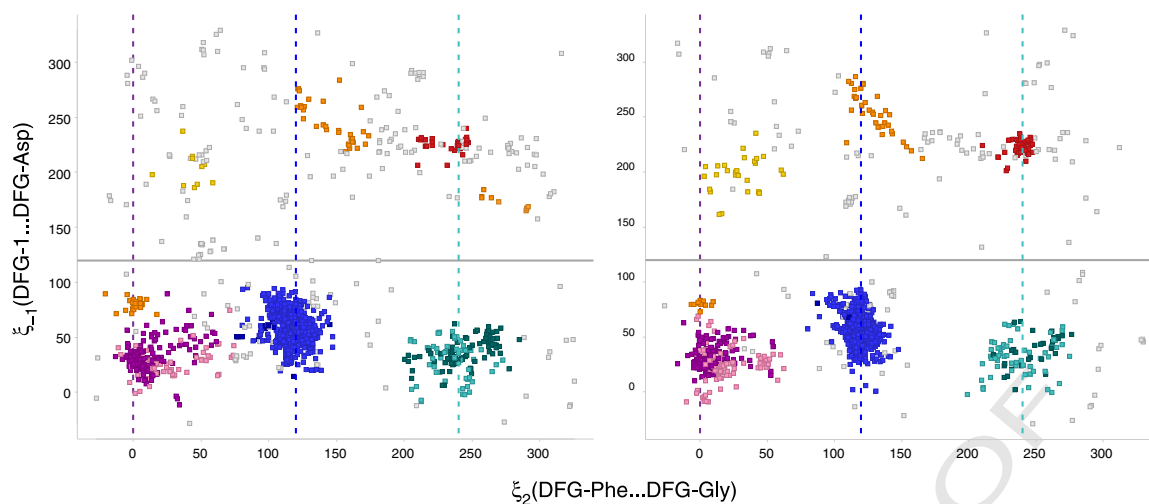


Fig. 1. DFG-plots of two sets of the PDB, post (left, 2171 chains) and pre June 2010 (right, 1909 chains), showing similar distributions of clusters. The main clusters are active (blue), FG-down (magenta) and G-down (cyan). DFG-out clusters are found above the border of $\xi_{-1} > 100$.

short review will briefly highlight some of these diverse approaches to computational kinase discovery presented at the conference.

2. Discussion

2.1. Conformation selection

A number of speakers described the interplay between active–inactive kinase conformations, and ways to computationally analyze and address them. Most kinase can be activated by phosphorylation of the activation loop, causing a conformational shift of the DFG motif from the “out” to the “in” positions, bringing the catalytic ASP into position to interact with phosphates on the ATP and magnesium ions to perform phosphate transfer. Henrik Moebitz presented a 3D alignment and structural clustering of all mammalian kinase X-ray conformations. The structures formed distinct clusters when plotted in 2 specialized graphs, a “DFG-plot” and a “Helix-C plot”, according to a few simple geometric criteria. The secret to getting distinct interpretable clusters was the identification of pseudo DFG torsions formed by sets of 4 consecutive alpha carbons, a measure of the torsion between two consecutive sidechains. These angles were divided into regions: FG-down/DFG-active/G-down, and DFG-in/out. The structures could then be plotted on the 2 graphs by adding a distance to helix-C, classified as in/dilated/out. Comparing two subsets of the PDB, prior and post June 2010, gave similar distributions of clusters (Fig. 1). Analyzing the populations

provided estimates of the energy differences between kinase conformational states. One interesting conclusion was that phosphorylation shifts the relative balance between active and inactive conformation by 1 kcal/mole on average (Fig. 2). This observation can explain why type II inhibitors, which bind in the inactive (DFG-out) conformation, exhibit lower potency (by 10 fold on average), but are measurable in biochemical (phosphorylated) kinase assays. He also observed that first-shell polar residues hinder the DFG transition. The research extends the long standing interest in classification of binding modes of kinase inhibitors [1].

Aiming to understand energetic and conformational preferences leading to observed selectivity, Benoit Roux and his co-workers Yen-Lin Lin and Yilin Meng described free energy molecular dynamic simulations to directly calculate the binding affinity of Imatinib, which inhibits Abl and c-Kit, but not c-Src, even though all three have >30% sequence homology. He drove 2 pseudo dihedrals by umbrella sampling to get full 2D conformational free energy maps of the unphosphorylated proteins around the DFG motif region of the activation loop. The maps showed only 2 stable conformations: DFG-in and DFG-out. According to the umbrella sampling calculations, Abl kinase appears to be more stable in the DFG-in conformation by a modest 1.4 kcal/mol, while Src is more stable in the DFG-in by 5.4 kcal/mol, suggesting that the free energy cost of the DFG flip between these two kinases could be one determinant of type II selectivity [2]. The team also calculated the affinity of Imatinib to the binding pocket by using the “alchemical double

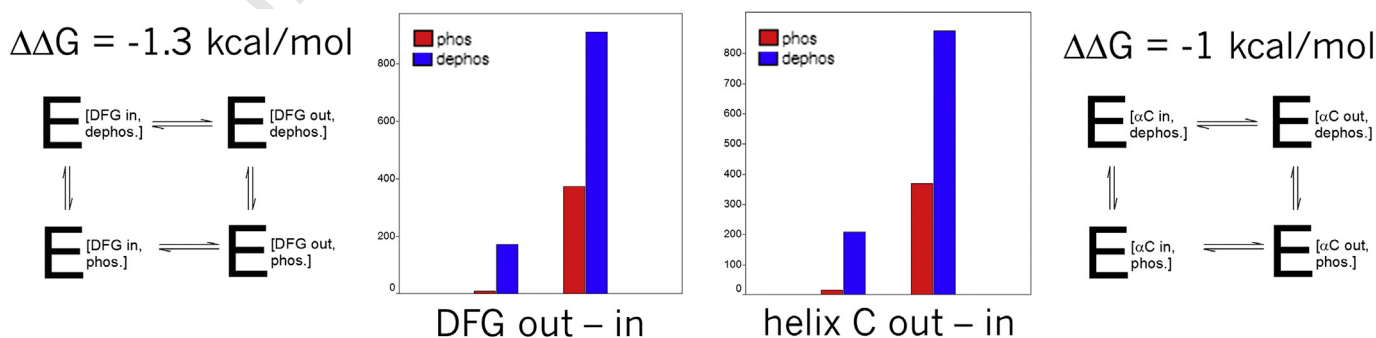


Fig. 2. Estimation of conformational bias from phosphorylation. Based on the population shift between DFG in and out conformations, the stabilizing effect of phosphorylation can be estimated from these thermodynamic cycles.

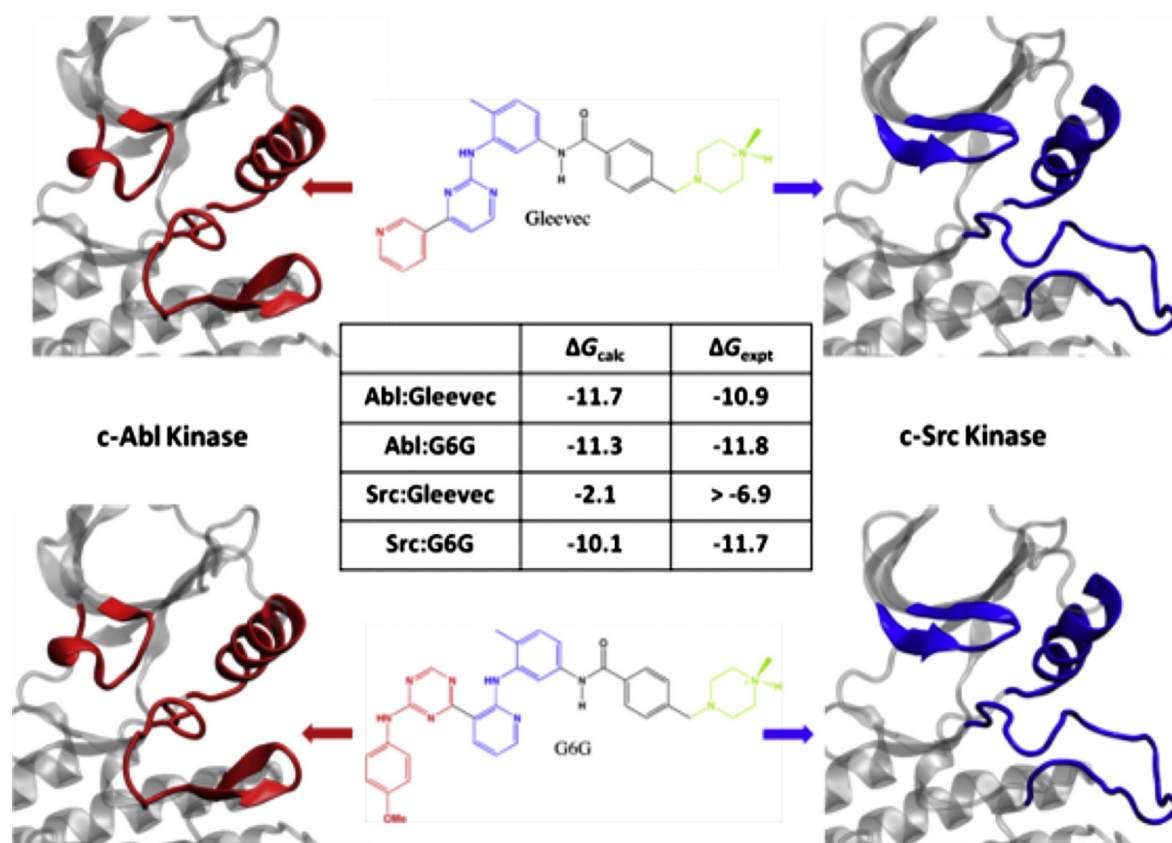


Fig. 3. Free energy differences from simulations agree well with experiments [2].

120 decoupling” technique to “annihilate” the inhibitor from solution and
 121 grow it into the protein. They decomposed the total free energy differ-
 122 ence, which agrees well with experiment (Fig. 3), into components by
 123 vanishing and growing the ligand one energy term at a time: first the re-
 124 pulsive part, then the van der Waals dispersion part, and finally the elec-
 125 trostatic part. This analysis, in close agreement with experimental data,
 126 correctly concluded that Imatinib binds Abl better, and identified the
 127 van der Waals dispersion term as the dominant energy component. In
 128 addition, the binding of an analog of Gleeevec (G6G), which is equally po-
 129 tent for Abl and Src, was investigated and agreement with experiment
 130 in binding affinity was observed [2]. This additional set of free energy
 131 calculations further supported that both conformational selection and
 132 protein–ligand interaction are responsible for the specificity of Gleeevec.

133 Stefan Knapp presented a wide range of experimental studies inves-
 134 tigating compounds which stabilize inactive conformations of kinases.

In collaboration with Nathanael Grays laboratory, he reported that
 135 more than 200 kinases, covering all branches of the kinome, were
 136 found to be inhibited by a small set of type 2 inhibitors, suggesting
 137 that a large fraction of kinases can be targeted by type II inhibitors [3].
 138 Type II inhibitor structures are underrepresented in the protein data
 139 based (PDB) and several type-II inhibitors were co-crystallized with ki-
 140 nases for which no experimental type-II structure has been reported.
 141 These structures included CDK2.

142 A unique binding mode was reported for the ERK inhibitor
 143 SCH772984, which bound in a so far unreported conformation to
 144 ERK1 and ERK2 (Fig. 4). In this novel binding mode, which would be im-
 145 possible to predict with current computational approaches, the inhibi-
 146 tor induced a binding pocket between the P-loop and αC , forming a
 147 number of hydrogen bonds and aromatic stacking interactions with resi-
 148 dues present in these structural elements. Binding of SCH772984 was
 149

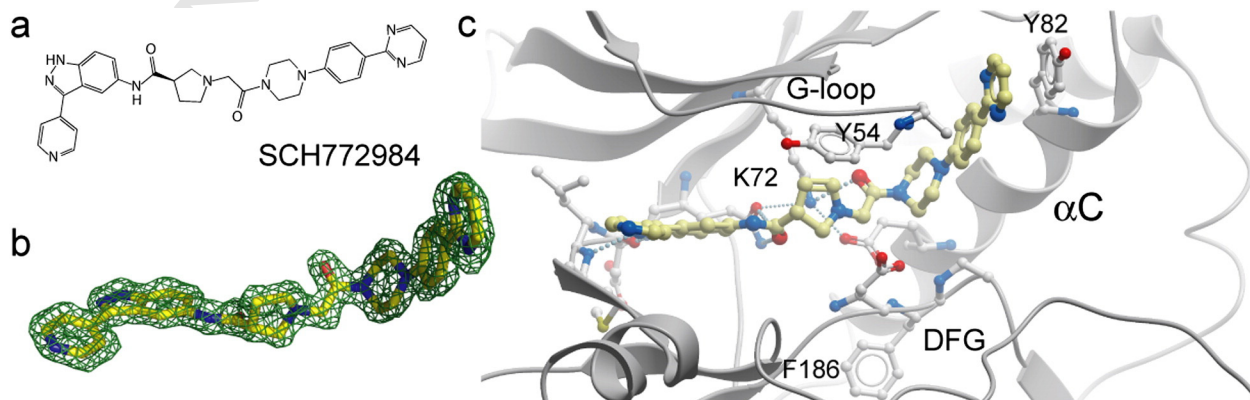


Fig. 4. Novel binding mode of SCH772984 in ERK2. a: Chemical structure of SCH772984. b: 2FoFc OMIT electron density map contoured at 2 σ . c: Details of the interaction in ERK2. C.

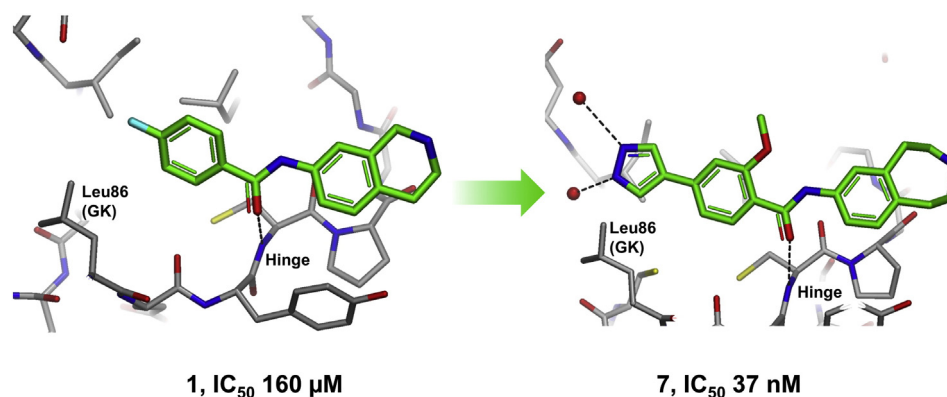


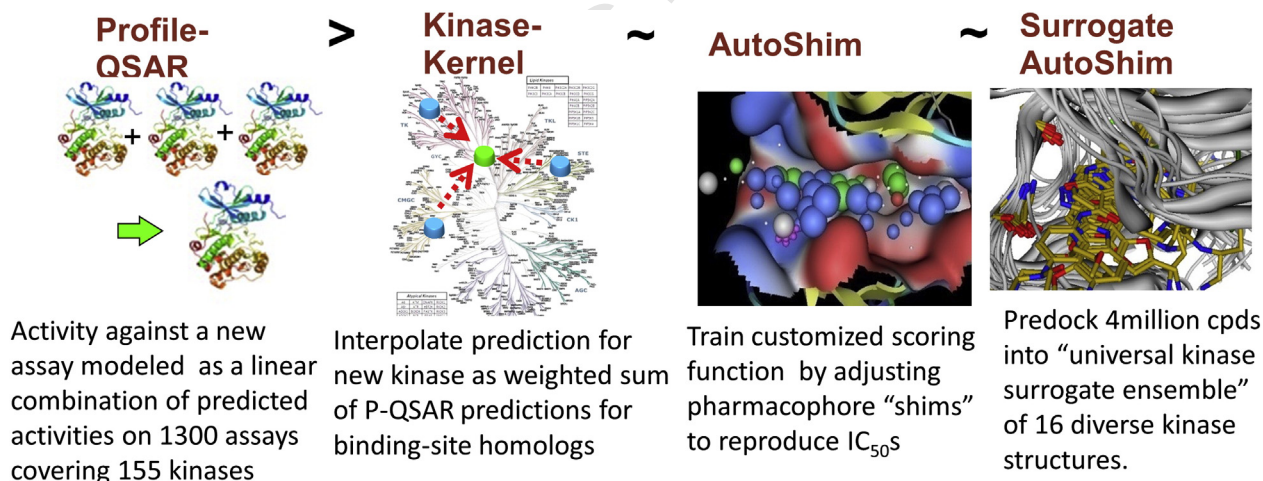
Fig. 5. A low affinity fragment hit was optimized by SBDD to a selective 37 nM tool compound.

150 associated with slow off rates *in vitro* as well as in cellular assays, where-
 151 as off-targets such as haspin and JNK interacted with the inhibitor in
 152 diverse but canonical type I binding modes and showed fast on and
 153 off-rates. Mutagenesis studies suggested that aromatic stacking interac-
 154 tions of residues located in α C, as well as the glycine rich loop, were im-
 155 portant for the slow binding kinetics of this inhibitor. The novel binding
 156 pocket may offer an alternative design strategy for type II inhibitors [4].

157 Valerio Berdini used MELK kinase as an example of how medicinal
 158 chemistry can use fragment starting points to create insights into stabi-
 159 lizing unique kinase conformations [5,6]. From 231 fragments that
 160 showed an effect in a protein melting-point screen, 144 confirmed in
 161 NMR. Subsequent X-ray crystallography showed 20 novel hinge
 162 binders. Isoquinoline fragments were optimized into both highly

163 efficient (LE = 0.54) type I ligands, and highly potent type II inhibitors. 163
 164 MELK has a large leucine gate keeper, and traditional type II linkers did
 165 not induce the DFG out conformation. On average, type I fragments and
 166 inhibitors had much higher ligand efficiencies, suggesting that the type
 167 II conformation in MELK is higher energy.

168 One of the Type I starting points was optimized into a selective Melk
 169 inhibitor that offered conformational selection for the MELK hinge re-
 170 gion [5]. A path from an initial, relatively inefficient 160 μ M fragment
 171 with unique binding, to the optimized 37 nM molecule with good
 172 selectivity, involved using a variety of structure based design tools and
 173 computational analog modeling to identify strong interactions with
 174 MELK (Fig. 5). Another approach utilizing the ASTEX structural
 175 informatics platform allowed for the rational design of a 19 nM type II



Method/ Feature	2D Profile-QSAR	2D Kinase-Kernel	3D (Plain) AutoShim	3D Surrogate AutoShim
Enzyme IC ₅₀	$R^2_{ext}=0.6$	$R^2_{ext}=0.5$	$R^2_{ext}=0.5$	$R^2_{ext}=0.5$
Selectivity	$R^2_{ext}=0.6$	☹	☹	☹
Cellular TM EC ₅₀	$R^2_{ext}=0.6$	☹	☹	☹
Structure free	☺	☺	☹	☺
Fast	☺	☺	☹	☺
IC ₅₀ data needed	☹	☺	☹	☹
Non-family target	☹	☹	☺	☹

Fig. 6. Four modeling methods comprising Protein-Family Virtual Screening.

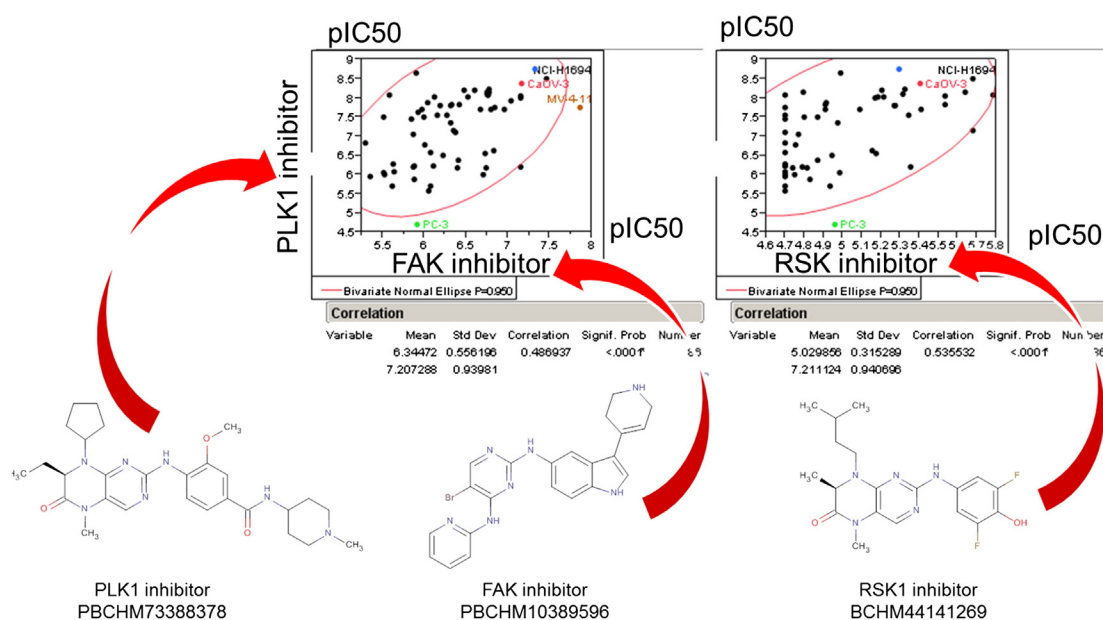


Fig. 7. Similarity of PLK1 (BI-2536), RSK (BI-D1870) and FAK (PBCHM10389596) inhibitors cancer cell sensitivity profiles. Whereas these three compounds were developed and optimized for different targets, they show a similar activity on a large cancer cell sensitivity panel.

176 inhibitor, although with a less optimal selectivity profile [6]. In this
 177 approach, existing structural fragments of hinge binders, linkers and
 178 positively ionizable groups were combined to stabilize the type II
 179 MELK conformation. Structure-based design was employed together
 180 with computational tools in the course of project evolution.

181 2.2. Predictive modeling

182 Predictive models are widely used for virtual screening against ki-
 183 nase targets. Both ligand-based, structure-based and mixed models
 184 are used in an industrial setting to initiate and focus kinase inhibitor
 185 discovery efforts. Kinome-wide profiling data allow the creation and
 186 evaluation of computational models not only for activity but also
 187 selectivity predictions. Thibault Varin presented an application of

188 ligand-based models in screening [7] campaigns at Lilly, and the discov-
 189 ery and initial optimization of selective RIO2 kinase inhibitors. Using
 190 chemical similarity, he selected from a set of virtual, robot-capable reac-
 191 tions a set of 8 compounds. These were robotically synthesized [8] and
 192 tested for activity [9]. Three showed activity improvement ranging
 193 from 2 to 10-fold from the initial hit.

194 Eric Martin described a collection of empirical protein-family virtual
 195 screening (PFVS) models (Fig. 6) which combine extensive IC_{50} and
 196 structural data from all historical kinase projects to produce predictive
 197 activity and selectivity models for both biochemical and cellular assays
 198 of new kinases, with accuracy comparable to experimental high-
 199 throughput screens [10]. He described numerous case studies where
 200 accurate prediction of biochemical and cellular selectivity identified
 201 starting points for medicinal chemistry and tool compounds that

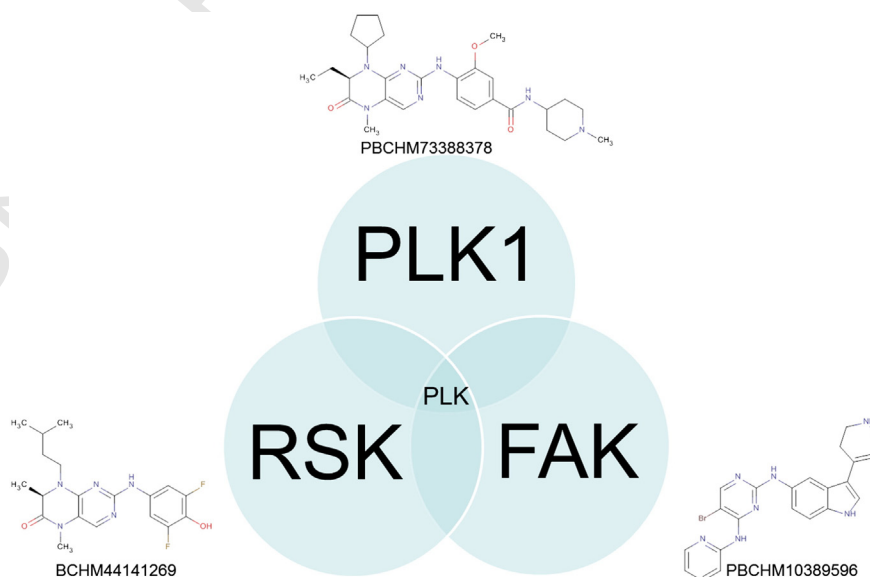


Fig. 8. The PLK1, RSK and FAK inhibitors are not fully selective and all inhibit also PLK1. As PLK1 is an essential gene, this could explain the similar activity of these compounds on the cancer cell sensitivity panel profile.

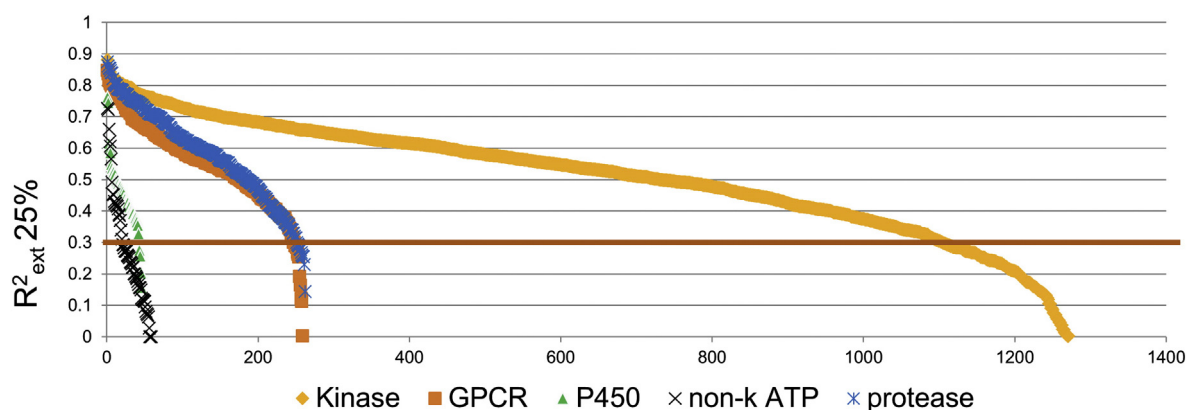


Fig. 9. IC_{50} prediction vs. experimental correlations for 25% held-out test sets for 2000 Profile-QSAR models covering 5 protein families. Predictions for 3,000,000 compounds have been pre-calculated and stored.

validated, or in several instances invalidated, newly proposed drug targets. Hit rates were consistently 25% to 80%, even for novel scaffolds completely unrelated to the known inhibitors.

2.3. Biochemical and cellular assay panels and computational target identification

Panels of biochemical and cellular assays [11] have been used in multiple ways to understand potential uses of kinase inhibitors to treat various cancers. The signatures together with Genetic backgrounds, mRNA expression levels and shRNA data have in turn been used to understand compound signatures, with the goal of creating patient tailoring hypotheses.

Thibault Varin showed how one could utilize kinase inhibitor profiles to elucidate reasons for cell panel signature similarity. In several cases, a target hypothesis could be derived, indicating and confirming the role of PLK1 [12] in cell proliferation. He presented comparisons of compounds based on the activity on large cancer cell sensitivity and kinase affinity panels. By integrating these two compound profiles, he showed that the target toward which a compound has been historically optimized doesn't necessarily drive the cellular activity of a given cell line or even of the overall cancer cell line panel. A RSK (BI-D1870) and a FAK (PBCHM10389596) inhibitor were reported to have a similar cancer cell sensitivity panel as a PLK1 (BI-2536) inhibitor. These compounds both

showed affinity for PLK1 in the kinase panels (Figs. 7 & 8) (Kinomescan, DiscoverX).

Eric Martin applied Protein Family Virtual Screening (PFVS) (see above) to predict the IC_{50} s for 3 million compounds against 2000 biochemical and cellular assays (Fig. 9) [7,13]. These were applied to predicting polypharmacology, modes-of-action for phenotypic screens, toxicity profiling, and selecting commercial compounds with diverse selectivity profiles for chemical archive enhancement.

Alexander Baumann described the extension of the well-established kinomescan methodology to the bromodomain (BRD) family. Screening kinase inhibitor libraries against a bromodomain panel identified several established kinase inhibitors that were cross-reactive and might be repurposed as kinase-BRD dual inhibitors. He also introduced the "BioMAP" technology platform that provides a measure of overall phenotypic response of compounds under disease-like conditions, and identifies clinically relevant activities across a broad protein biomarker panel. The BioMAP systems are stimulated primary human cell types and co-cultures designed to recapitulate the complex signaling networks and microenvironment in diseased human tissue (Fig. 10) [14]. The resulting biomarker fingerprints are useful for identifying modes of action and to toxicity profiling. The biomarker fingerprints from the kinase/BRD dual inhibitors showed a hybrid of both mechanisms [15].

Richard Engh examined methods to evaluate protein kinase target similarities with the aim to compare information types hierarchically. At the simplest level, "pseudosequence" similarities were calculated

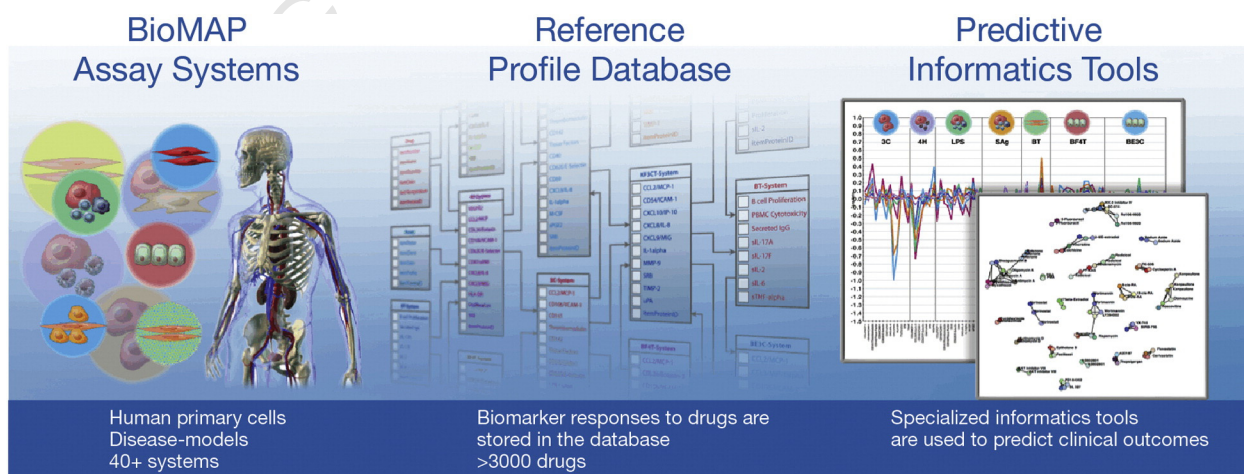


Fig. 10. The BioMAP system showing Human Primary cells Disease models on the left, biomarker responses to >3000 drugs stored in the database in the middle and a variety of specialized informatics tools with ability to predict clinical outcomes from data.

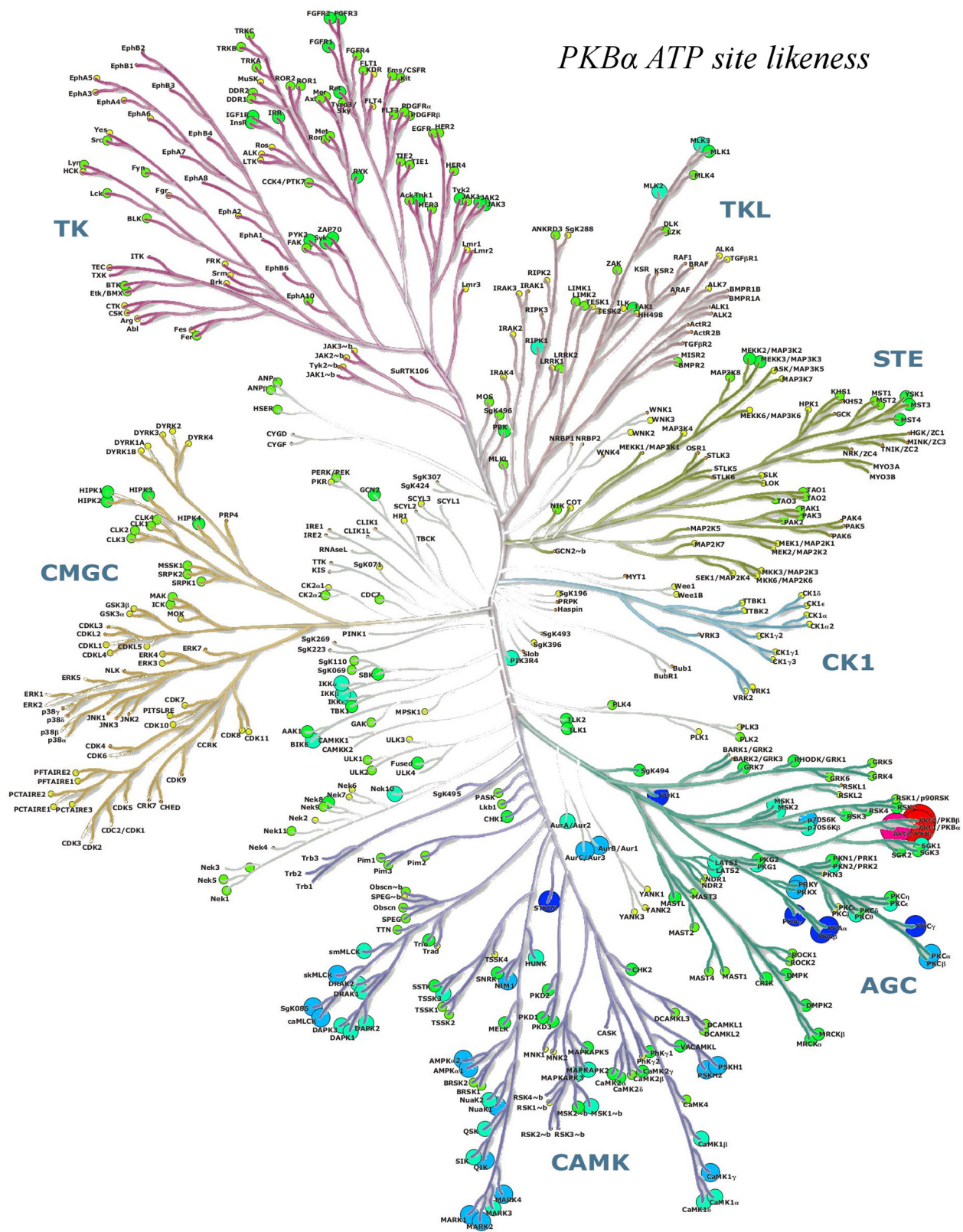


Fig. 11. Disk sizes and colors depict pseudosequence similarities to PKB alpha.

249 based on sequences chosen to represent binding site residues (Fig. 11).
 250 Statistically, these corresponded quite well with experimental inhibition
 251 profiles from Ambit 2011 data [16], especially for tyrosine kinase
 252 targets (Fig. 12). Such analyses support the use of surrogate kinases in

structure-based drug discovery [17], and may aid in choosing focused
 screening libraries for repurposing or retargeting known compounds.

254 At a higher level of information content, similarity analyses of target
 255 proteins would involve comparisons of X-ray structures. Currently, 256

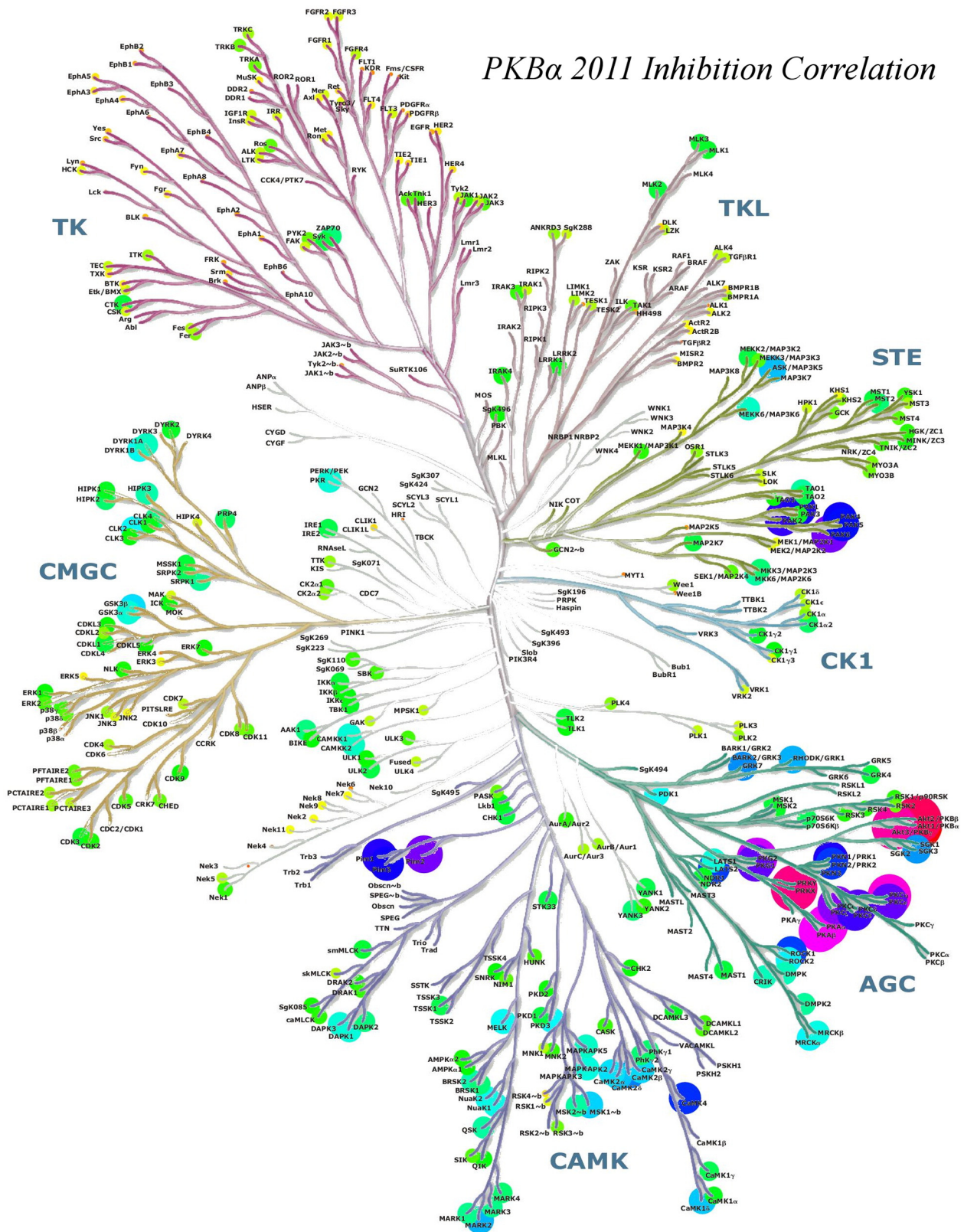


Fig. 12. Disk sizes and colors depict correlation of AMBIT 2011 inhibition profiles of protein kinase targets with those of PKB alpha. Note similarities to Fig. 11, pseudosequence similarities.

comprehensive integration of structural information is not practically possible: there is too much unpredictable structural variability, the distributions of structural states are strongly influenced by crystallization conditions [18], and experimental binding data are highly dependent

on assay conditions, which in turn are often not accessible to data mining tools. On the other hand, some specific areas are well supported, including reliable clustering of key structural states (notably DFG and C-helix states, see many other talks from IPK2014 and other references, 264

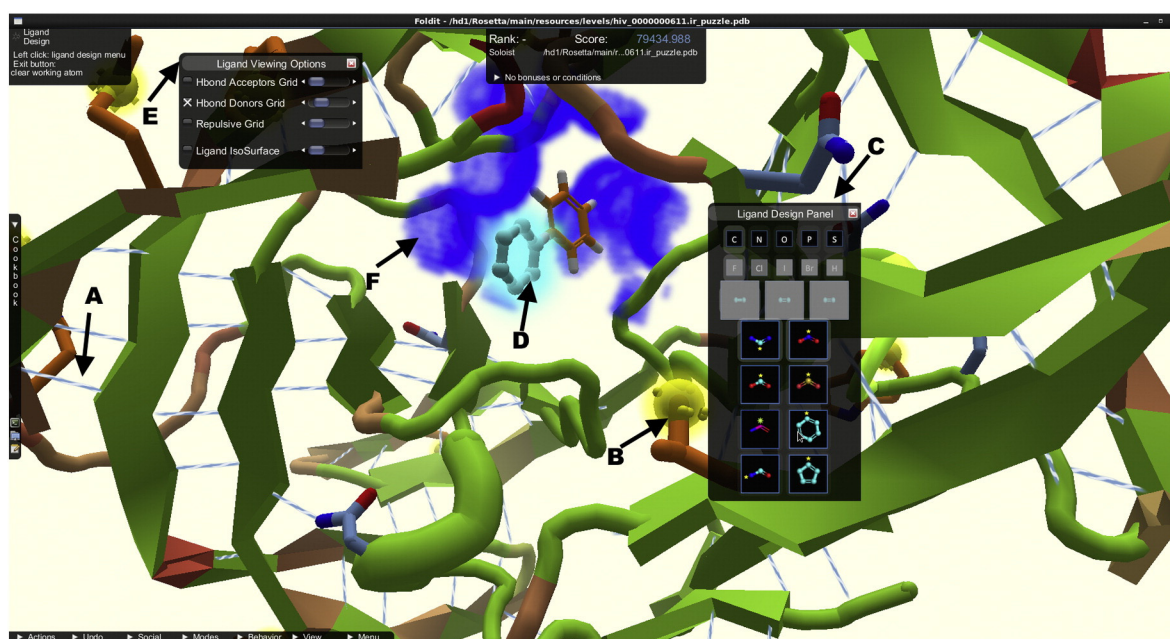


Fig. 13. Screenshot of Foldit, from the new Rosetta Ligand application, a crowd-sourced multiplayer game adopted for ligand design. A) Hydrogen bond contacts are shown in light-blue/white lines and B) surface exposed hydrophobic residues are shown as yellow blobs. C) The ligand design panel is the control center for players and allows them to choose from a variety of fragments, bond manipulations, and element modifications to design the ligand in the protein binding pocket. D) When players hover over a fragment, a ghost view of the new fragment is drawn at the attachment point (light blue glowing fragment). E) The ligand viewing option menu allows players to turn off QSAR grids calculated for hydrogen bond acceptors and donors (shown as dark blue fog) or repulsion for the ligand, with slider bars on the side to adjust the alpha of the drawn fogs. Players can turn on the protein isosurface or a ligand centric view that draws the isosurface around the ligand, allowing for advance spatial alignment of the ligand in the binding pocket.

265 including [19]). For well characterized diseases, the data increasingly
266 enable targeted pharmacology by identifying key determinants of target
267 similarities, possibly combined with “orthogonal” dissimilarities.

268 2.4. Future directions—introducing crowd sourcing into drug design

269 Jens Meiler showed principles and application examples of the
270 RosettaLigand [20] and BCL::Cheminformatics [21] computational soft-
271 ware packages, developed across academic institutions, highlighting
272 both structure-based and ligand-based drug design (Fig. 13). A fascinat-
273 ing application of Rosetta is the computer game Foldit [22], with over
274 200,000 users. The goal of the game is to predict the structure of a pro-
275 tein. In addition to educational value, it is an example of crowd sourcing
276 to solve challenging scientific problems. The crowd sourcing approach is
277 similarly being translated into drug discovery with a drug design
278 component of Foldit. One application of the game was to one of
279 malaria’s essential kinases PKG [23], the X-ray structure of which was
280 revealed at the meeting. For ligand-based drug design the QSAR applica-
281 tion BCL::Cheminformatics [21] was introduced which converted a
282 ligand structure into a property vector of charge, shape, or H-bonds
283 donors and acceptors. Chirality was included by a signed volume using
284 a right-hand rule [24]. A QSAR model was trained using an artificial neu-
285 ral network. In several example applications the hit-rate in virtual
286 screening increased by factors of 15–50 over conventional diversity
287 approaches.

288 3. Conclusions and summary

289 These computational presentations illustrated many recent ad-
290 vances in understanding the activity and selectivity of kinase inhibitors
291 and their relationships to protein structure. The approaches ranged
292 from highly empirical to purely physics-based. They ranged from
293 mining large activity and structure databases, to simulating the physics
294 of ligand binding into active and inactive conformations, to probing

conformational energetics by synthesizing related ligands designed to
295 bind alternate conformations. The presentations helped to move our
296 understanding of the chemistry, physics, biochemistry and biology of
297 kinase structure and activity, and illustrated how that understanding
298 impacts drug discovery programs. It is also our view that computational
299 methods will facilitate drug discovery/development against kinases of
300 eukaryotic pathogens for which there is limited experimental informa-
301 tion available, an additional theme of the conference. Hopefully these
302 highlights have whetted your appetite to dig further into some of
303 these topics in the accompanying articles in this issue.
304

305 Conflict of interests

306 The authors declare having no conflict of interest.

307 Acknowledgments

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309 for their continued support of this conference series and Dr. Steven
310 Combs for his contributions and research on drug design in Foldit.

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